



## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

<b>(51) International Patent Classification <sup>6</sup> :</b> <b>C12N 15/85, 5/10, C12Q 1/42, 1/48,</b> <b>C07K 14/72 // C12N 9/16</b>	<b>A1</b>	<b>(11) International Publication Number:</b> <b>WO 95/23231</b> <b>(43) International Publication Date:</b> 31 August 1995 (31.08.95)
<b>(21) International Application Number:</b> PCT/EP95/00731 <b>(22) International Filing Date:</b> 28 February 1995 (28.02.95) <b>(30) Priority Data:</b> 08/203,218 28 February 1994 (28.02.94) US <b>(71) Applicant:</b> MAX-PLANCK-GESELLSCHAFT ZUR FÖRDERUNG DER WISSENSCHAFTEN E.V. [DE/DE]; Hofgartenstrasse 2, D-80539 München (DE). <b>(72) Inventors:</b> ULLRICH, Axel; Adalbertstrasse 108, D-80798 München (DE). HOPPE, Edmund; Bergstrasse 2b, D-82152 Krailling (DE). MOLLER, Neils, P., H.; Heiterwanger Strasse 32, D-81373 München (DE). <b>(74) Agents:</b> RUFF, Michael et al.; Willy-Brandt-Strasse 28, D- 70173 Stuttgart (DE).		<b>(81) Designated States:</b> AM, AT, AU, BB, BG, BR, BY, CA, CH, CN, CZ, DE, DK, EE, ES, FI, GB, GE, HU, JP, KE, KG, KP, KR, KZ, LK, LR, LT, LU, LV, MD, MG, MN, MW, MX, NL, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TT, UA, UZ, VN, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG), ARIPO patent (KE, MW, SD, SZ, UG).  <b>Published</b> <i>With international search report.</i> <i>Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>
<b>(54) Title:</b> CELL LINES FOR THE IDENTIFICATION OF SUBSTANCES AFFECTING INSULIN RECEPTOR MEDIATED SIGNAL TRANSDUCTION  <b>(57) Abstract</b>  The present invention relates to cell lines useful for the screening and identification of compounds that by modulating phosphotyrosine phosphatase activity, modulate insulin receptor type tyrosine kinase mediated signal transduction. Genetically engineered cells expressing IR in culture overcome the effect of insulin on morphology and adhesion when they are also coexpressing RPTP $\alpha$ or RPTP $\epsilon$ . Such engineered cell lines may be used to screen and identify non-toxic compounds that could elicit or modulate insulin signal transduction even in the absence of insulin.		

**FOR THE PURPOSES OF INFORMATION ONLY**

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AT	Austria	GB	United Kingdom	MR	Mauritania
AU	Australia	GE	Georgia	MW	Malawi
BB	Barbados	GN	Guinea	NE	Niger
BE	Belgium	GR	Greece	NL	Netherlands
BF	Burkina Faso	HU	Hungary	NO	Norway
BG	Bulgaria	IE	Ireland	NZ	New Zealand
BJ	Benin	IT	Italy	PL	Poland
BR	Brazil	JP	Japan	PT	Portugal
BY	Belarus	KE	Kenya	RO	Romania
CA	Canada	KG	Kyrgystan	RU	Russian Federation
CF	Central African Republic	KP	Democratic People's Republic of Korea	SD	Sudan
CG	Congo	KR	Republic of Korea	SE	Sweden
CH	Switzerland	KZ	Kazakhstan	SI	Slovenia
CI	Côte d'Ivoire	LI	Liechtenstein	SK	Slovakia
CM	Cameroon	LK	Sri Lanka	SN	Senegal
CN	China	LU	Luxembourg	TD	Chad
CS	Czechoslovakia	LV	Latvia	TG	Togo
CZ	Czech Republic	MC	Monaco	TJ	Tajikistan
DE	Germany	MD	Republic of Moldova	TT	Trinidad and Tobago
DK	Denmark	MG	Madagascar	UA	Ukraine
ES	Spain	ML	Mali	US	United States of America
FI	Finland	MN	Mongolia	UZ	Uzbekistan
FR	France			VN	Viet Nam
GA	Gabon				

CELL LINES FOR THE IDENTIFICATION OF SUBSTANCES  
AFFECTING INSULIN RECEPTOR MEDIATED SIGNAL  
TRANSDUCTION

5

1. INTRODUCTION

The present invention relates to genetically  
engineered cells useful for the screening and  
10 identifying of compounds that affect insulin receptor-  
type tyrosine kinase mediated signal transduction.

The present invention further relates to methods  
for screening and identifying of specific compounds,  
that by modulating the activity of the controlling  
15 protein phosphotyrosine phosphatases, have uses in the  
treatment of diabetes and other diseases.

2. BACKGROUND OF THE INVENTION

20

2.1 SIGNAL TRANSDUCTION

Cellular signal transduction is a fundamental  
mechanism whereby external stimuli regulate diverse  
cellular processes are relayed to the interior of  
25 cells. The process is generally initiated by the  
binding of extracellular factors (such as hormones and  
growth factors) to membrane receptors on the cell  
surface. The biochemical pathways through which  
signals are transmitted within cells comprise a  
30 circuitry of directly or functionally connected  
interactive proteins.

One of the key biochemical mechanisms of signal  
transduction involves the reversible phosphorylation  
of tyrosine residues on proteins. The phosphorylation  
35 state of a protein may affect its conformation and/or  
enzymic activity as well as its cellular location.

- 2 -

The phosphorylation state of a protein is modified through the reciprocal actions of protein tyrosine kinases (PTKs) and protein phosphotyrosine phosphatases (PTPs). Generally, the level of tyrosine phosphorylation increases after the cell has been stimulated by an extracellular factor. Research has largely focussed on the protein kinases (Sefton et al., 1980, Cell 20:807-16; Heldin and Westermark, 1984, Cell 37:9-20; Yarden and Ullrich, 1988, Ann. Rev. Biochem. 57:443-78; Ullrich and Schlessinger, 1990, Cell, 61:203-12).

Protein tyrosine kinases comprise a large family of transmembrane as well as cytoplasmic enzymes with multiple functional domains (Taylor et al., 1992, Ann. Rev. Cell Biol. 8:429-62). The binding of an extracellular factor or ligand allosterically transduces the signal to the inner face of the cell membrane where the cytoplasmic portion of the receptor protein tyrosine kinase (RPTKs) initiates a cascade of molecular interactions that disseminate the signal throughout the cell and into the nucleus.

Ligand-induced activation of the kinase domain and its signalling potential are mediated by receptor dimerization. Once activated, the receptor self-phosphorylates (autophosphorylation or transphosphorylation) on specific tyrosine residues of the cytoplasmic domain. (Schlessinger, 1988, Trends Biochem. Sci. 13:443-7, Schlessinger and Ullrich, 1992, Neuron, 9:383-91, and references therein).

Like the PTKs, the protein phosphotyrosine phosphatases (PTP) comprise a family of transmembrane and cytoplasmic enzymes. (Hunter, 1989, Cell, 58:1013-16; Fischer et al., 1991, Science, 253:401-6; Saito and Streuli, 1991, Cell growth and differentiation, 2:59-65; Pot and Dixon, 1992,

- 3 -

Biochim. Biophys. Acta, 1136:35-43). As presently understood by those in the art, in general PTKs play a triggering role in signal transduction, while PTPs  
5 guarantee that the trigger is reset thereby serving to deactivate the pathway. However, the specific functions of PTPs have not yet been defined (Walton et al., 1993, Ann. Rev. Biochem., 66:101-20).

In addition to a homologous core catalytic  
10 domain, mammalian PTPs share diverse noncatalytic sequences. While some receptor protein tyrosine phosphatases (RPTPs) contain in their extracellular portions Ig-like and/or fibronectin type III repeats (e.g., LAR, Streuli et al., 1988, J. Exp. Med.  
15 168:1523); others have small extracellular glycosylated segments (e.g., RPTP $\alpha$ , Sap et al., 1990, Proc. Natl. Acad. Sci. USA, 87:6112; and RPTP $\epsilon$ , Krueger et al., 1990, EMBO J, 9:3241). In all cases, the putative ligands have yet to be identified. Other  
20 phosphotyrosine phosphatases such as PTP1B, PTP $\mu$ , PTP1C, TC-PTP, PTPH1, RPTP $\kappa$  and CD45 have been cloned and their cDNAs are described in Chernoff et al., 1990, Proc. Natl. Acad. Sci. USA, 87:2735-9; Gebbink et al., 1991, FEBS Lett. 290:123-30; Shen et al.,  
25 1991, Nature, 352:736-9; Jiang et al., 1993, Mol. Cell Biol., 13:2942-51 and; Charbonneau et al., 1988, Proc. Natl. Acad. Sci. USA, 85:7182-6 respectively.

Abnormal PTK/PTP signal transduction has been associated with a variety of diseases including  
30 psoriasis, cancer and diabetes.

## 2.2 THE INSULIN RECEPTOR AND DIABETES MELLITUS

35 The insulin receptor (IR) (Ullrich et al., Nature, 313:756-61, 1985) is the prototype for a family of RPTKs structurally defined as a heterotetrameric

species of two  $\alpha$  and two  $\beta$  subunits. Other members of the insulin receptor-type protein tyrosine kinase (IR-PTK) family include, for example, the receptor for  
5 insulin-like growth factor 1 (IGF-1 R, Ullrich et al., 1986, EMBO J. 5:2503-12) and insulin related receptor (IRR, Zhang et al., 1992, J. Biol. Chem. 267:18320-8) the ligand(s) for which is at present unknown.

The binding of insulin to the insulin receptor  
10 triggers a variety of metabolic and growth promoting effects. Metabolic effects include glucose transport, biosynthesis of glycogen and fats, inhibition of triglyceride breakdown, and growth promoting effects include DNA synthesis, cell division and  
15 differentiation. It is known that some of these biological effects of insulin can be mimicked by vanadium salts such as vanadates and pervanadates. However, this class of compounds appears to inhibit phosphotyrosine phosphatases generally, and are  
20 potentially toxic because they contain heavy metal (U.S. Patent No. 5,155,031; Fantus et al., 1989, Biochem., 28:8864-71; Swarup et al., 1982, Biochem. Biophys. Res. Commun. 107:1104-9).

Diabetes mellitus is a heterogeneous primary  
25 disorder of carbohydrate metabolism with multiple etiologic factors that generally involve insulin deficiency or insulin resistance or both. Type I, or juvenile onset, or insulin-dependent diabetes mellitus, is present in patients with little or no  
30 endogenous insulin secretory capacity. These patients develop extreme hyperglycemia and are entirely dependent on exogenous insulin therapy for immediate survival. Type II, or adult onset, or non-insulin-dependent diabetes mellitus, occurs in patients who  
35 retain some endogenous insulin secretory capacity but the great majority of them are both insulin deficient

- 5 -

and insulin resistant. Insulin resistance can be due to insufficient insulin receptor expression, reduced insulin-binding affinity, or any abnormality at any  
5 step along the insulin signaling pathway (Olefsky, 1988, in "Cecil Textbook of Medicine," 18th Ed., 2:1360-81)

Overall, in the United States the prevalence of diabetes is probably between 2 and 4 per cent, with  
10 Type I comprising 7 to 10 per cent of all cases. Secondary complications of diabetes have serious clinical implications, such as amputations (primarily of toes, feet, and legs) and blindness.

Insulin is the primary mode of therapy in all  
15 patients with Type I diabetes and in many with Type II diabetes. Oral hypoglycemic agents such as sulfonylureas are effective in Type II diabetic patients but approximately 10 to 20 per cent of patients do not respond or cease to respond 12-24  
20 months after treatment began.

Effective control of glucose level is difficult to achieve for prolonged periods even with the most meticulous mode of insulin therapy in the most motivated patients. Transplantation of the pancreas  
25 or islet cells, which normally produce insulin, continues to receive extensive study as a potential treatment. In addition, efforts towards developing newer and better external or implantable insulin-delivery devices integrated with a glucose sensor  
30 continues.

### 3. SUMMARY OF THE INVENTION

The present invention relates to cell lines  
35 useful for the screening and identification of

- 6 -

compounds that modulate insulin receptor-type tyrosine kinase (IR-PTK) mediated signal transduction.

The invention is based, in part, on the discovery  
5 that genetically engineered cells coexpressing IR and RPTP $\alpha$  or RPTP $\epsilon$  in culture are not sensitive to the effects of insulin on cell morphology and adhesion. The phenotype of the cells may be used as an indicator of insulin mediated signal transduction. The claimed  
10 cell lines of the invention are, therefore, useful in screening assays for non-toxic compounds, that by modulating phosphatase activity, modulate or prolong IR-PTK signal transduction.

In specific embodiments of the present invention  
15 detailed in the example section *infra*, the stable coexpression of IR and RPTP $\alpha$  or RPTP $\epsilon$  in baby hamster kidney (BHK) cells, and the development of cell-based assay system for IR signal transduction are described.

#### 20 4. BRIEF DESCRIPTION OF THE FIGURES

Figure 1A is a photograph showing the insulin-induced change in phenotype of a BHK cell line expressing the insulin receptor.

25 Figure 1B is a photograph showing the phenotype of a BHK cell line coexpressing the insulin receptor and RPTP $\alpha$  in the presence of insulin.

Figure 2A shows the phosphorylation status of IR in the presence or absence of insulin in two BHK cell  
30 clones transfected with the RPTP $\alpha$  gene: control expressing IR alone, clones 4 and 5 coexpressing IR and RPTP $\alpha$ . The filter was probed with anti-phosphotyrosine (anti-PY) antibodies. The molecular weight in kD is indicated.

35 Figure 2B shows the level of RPTP $\alpha$  expression in the presence or absence of insulin in BHK cell clones:



- 7 -

control expressing IR alone, clones 4 and 5  
coexpressing IR and RPTP $\alpha$ . The filter was probed with  
an anti-RPTP $\alpha$  antibody. The molecular weight in kD is  
5 indicated.

Figure 2C shows the level of IR expression in the  
presence or absence of insulin in BHK cell clones:  
control expressing IR alone, clones 4 and 5  
coexpressing IR and RPTP $\alpha$ . The filter was probed with  
10 an anti-IR antibody. The molecular weight in kD is  
indicated.

Figure 3A shows the phosphorylation status of IR  
in the presence or absence of insulin in BHK cell  
clones: control expressing IR alone, clones 4, 5 and 6  
15 coexpressing IR and RPTP $\epsilon$ . The filter was probed with  
anti-phosphotyrosine (anti-PY) antibodies. The  
molecular weight in kD is indicated.

Figure 3B shows the level of RPTP $\epsilon$  expression in  
the presence or absence of insulin in BHK cell clones:  
20 control expressing IR alone, clones 4, 5 and 6  
coexpressing IR and RPTP $\epsilon$ . The filter was probed with  
an anti-RPTP $\epsilon$  antibody.

Figure 3C shows the level of IR expression in the  
presence or absence of insulin in BHK cell clones:  
25 control expressing IR alone, clones 4, 5 and 6  
coexpressing IR and RPTP $\epsilon$ . The filter was probed with  
an anti-IR antibody. The molecular weight in kD is  
indicated.

30 5. DETAILED DESCRIPTION OF THE INVENTION

The present invention is directed to cell lines  
useful in screening assays for compounds that modulate  
insulin receptor-type tyrosine kinase (IR-PTK)  
35 mediated signal transduction. The IR-PTKs include  
insulin receptor, insulin-like growth factor 1 (IGF-1

- 8 -

R) and insulin receptor related receptor (IRR). As used herein, the term signal transduction is not limited to transmembrane signalling, and includes the multiple pathways that branch off throughout the cell and into the nucleus. The term ligand is synonymous with extracellular signalling molecules, and includes insulin, IGF-1, IGF-2 and other hormones, growth factors or cytokines that may interact with IR-PTKs.

Genetically engineered cells expressing IR are sensitive to the presence of insulin in culture and this sensitivity is easily detected. More specifically, the cells respond to insulin by losing their normal flat and adherent phenotype, and instead, round up and become detached from the culture dish. However, when these IR-expressing cells are transfected with DNA encoding RPTP $\alpha$  or RPTP $\epsilon$ , the cells coexpressing IR and the phosphatase are able to grow normally in the presence of insulin. Although, the inventors do not want to be bound by any specific theoretical mechanism, it is possible that the presence of the phosphatase restores balance to the signal transduction pathways activated by the insulin receptor in the presence of its ligand.

In a preferred embodiment of the invention, genetically engineered cell lines coexpressing IR and RPTP $\alpha$  or RPTP $\epsilon$  may be used to screen and identify compounds which, by modulating the activity of RPTP $\alpha$  or RPTP $\epsilon$ , elicit, modulate or prolong insulin receptor signal transduction.

#### 5.1 COEXPRESSION OF RPTPS AND IR-PTK AND GENERATION OF ENGINEERED CELL LINES

In accordance with the invention, RPTP $\alpha$ , RPTP $\epsilon$  and IR nucleotide sequences or functional equivalents thereof may be used to generate recombinant DNA

- 9 -

molecules that direct the coexpression of RPTP $\alpha$  or RPTP $\epsilon$  and IR proteins or a functionally equivalent thereof, in appropriate host cells. The nucleotide sequences of RPTP $\alpha$ , RPTP $\epsilon$  and IR are reported in Sap et al., 1990, Proc. Natl. Acad. Sci. USA, 87:6112-6 and Kaplan et al., 1990, Proc. Natl. Acad. Sci. USA, 87:7000-4; Krueger et al., 1990, EMBO J, 9:3241-52; and Ullrich et al., 1985, Nature 313:756-61 respectively and are incorporated by reference herein in their entirety. The specific interaction between RPTP $\alpha$ , RPTP $\epsilon$  and IR may involve the formation of a transient or stable multimolecular complex, hereinafter, referred to as RPTP $\alpha$ -IR, RPTP $\epsilon$ -IR complex or generally RPTP-IR-PTK complex. As used herein, a functionally equivalent RPTP $\alpha$ , RPTP $\epsilon$  or IR refers to an enzyme with essentially the same catalytic function, but not necessarily the same catalytic activity as its native counterpart. A functionally equivalent receptor refers to a receptor which binds to its cognate ligand, but not necessarily with the same binding affinity of its counterpart native receptor.

Due to the inherent degeneracy of the genetic code, other DNA sequences which encode substantially the same or a functionally equivalent amino acid sequence, may be used in the practice of the invention for the coexpression of the RPTP $\alpha$  or RPTP $\epsilon$  and IR proteins. Altered DNA sequences which may be used in accordance with the invention include deletions, additions or substitutions. For example, mutations may be introduced using techniques which are well known in the art, e.g. site directed mutagenesis, to insert new restriction sites, to alter glycosylation patterns, phosphorylation, etc. Amino acid substitutions may be made on the basis of similarity in polarity, charge, solubility, hydrophobicity,

- 10 -

hydrophilicity, and/or the amphipatic nature of the residues involved.

The RPTP $\alpha$ , RPTP $\epsilon$ , or IR or a modified RPTP $\alpha$ , RPTP $\epsilon$  or IR sequence may be ligated to a heterologous sequence to encode a fusion protein. For example, for screening of peptide libraries it may be useful to encode a chimeric RPTP $\alpha$ , RPTP $\epsilon$  or IR protein expressing a heterologous epitope that is recognized by an antibody. A fusion protein may also be engineered to contain the ligand-binding, regulatory or catalytic domain of another PTP or PTK.

The coding sequence of RPTP $\alpha$ , RPTP $\epsilon$  or IR could be synthesized in whole or in part, using chemical methods well known in the art. See, for example, Caruthers, et al., 1980, Nuc. Acids Res. Symp. Ser. 7:215-233; Crea and Horn, 180, Nucleic Acids Res. 9(10):2331; Matteucci and Caruthers, 1980, Tetrahedron Letters 21:719; and Chow and Kempe, 1981, Nucleic Acids Res. 9(12):2807-2817.

In order to coexpress a biologically active RPTP $\alpha$ , RPTP $\epsilon$  or IR, the nucleotide sequence coding for RPTP $\alpha$ , RPTP $\epsilon$  or IR, or their functional equivalent(s) as described supra, is inserted into one or more appropriate expression vector(s), i.e., a vector which contains the necessary elements for the transcription and translation of the inserted coding sequence(s). The RPTP $\alpha$  and/or RPTP $\epsilon$  gene(s) may be placed in tandem with the IR sequence under the control of the same or different promoter used to control the expression of the other coding sequence. The two phosphatases, RPTP $\alpha$  and RPTP $\epsilon$  may also be coexpressed together with IR.

Methods which are well known to those skilled in the art can be used to construct expression vectors containing the RPTP $\alpha$ , RPTP $\epsilon$  and/or IR coding sequence(s) and appropriate transcriptional/

- 11 -

translational control signals. These methods include in vitro recombinant DNA techniques, synthetic techniques and in vivo recombination/genetic recombination. See, for example, the techniques described in Maniatis et al., 1989, Molecular Cloning A Laboratory Manual, Cold Spring Harbor Laboratory, N.Y. and Ausubel et al., 1989, Current Protocols in Molecular Biology, Greene Publishing Associates and Wiley Interscience, N.Y.

A variety of host-expression vector systems may be utilized to coexpress the RPTP $\alpha$ , RPTP $\epsilon$ , or IR coding sequences. These include but are not limited to microorganisms such as bacteria transformed with recombinant bacteriophage DNA, plasmid DNA or cosmid DNA expression vectors containing the RPTP $\alpha$ , RPTP $\epsilon$ , or IR coding sequence(s) (see, Current Protocols in Molecular Biology, Vol. 2, 1988, Ed. Ausubel et al., Greene Publish. Assoc. & Wiley Interscience, Section 16.1); yeast transformed with recombinant yeast expression vectors containing the RPTP $\alpha$ , RPTP $\epsilon$ , or IR coding sequence(s) (Bitner, 1987, Heterologous Gene Expression in Yeast, Methods Enzymol, Eds. Berger & Mimmel, Acad. Press, N.Y. 152:673-84); insect cell systems infected with recombinant virus expression vectors (e.g., baculovirus, see Smith et al., 1983, J. Virol. 46:584; Smith, U.S. Patent No. 4,215,051) containing the RPTP $\alpha$ , RPTP $\epsilon$  and/or IR coding sequence(s); plant cell systems infected with recombinant virus expression vectors (e.g., cauliflower mosaic virus, CaMV; tobacco mosaic virus, TMV) or transformed with recombinant plasmid expression vectors (e.g., Ti plasmid) containing the RPTP $\alpha$ , RPTP $\epsilon$  and/or IR coding sequence(s) (see Weissbach & Weissbach, 1988, Methods for Plant Molecular Biology, Academic Press, NY); or animal cell systems.

- 12 -

In mammalian host cells, a number of viral based expression systems may be utilized. (E.g., See Logan & Shenk, 1984, Proc. Natl. Acad. Sci. (USA) 81:3655-3659, Mackett et al., 1982, Proc. Natl. Acad. Sci. (USA) 79:7415-7419; Mackett et al., 1984, J. Virol. 49:857-864).

A host cell of a particular cell type may also be chosen for the cell type-specific cofactors which may be required for the specific signalling pathway. A host cell strain may also be chosen which modulates the expression of the inserted sequences, or modifies and processes the gene product in the specific fashion desired. Such modifications (e.g., glycosylation) and processing (e.g., cleavage) of protein products may be important for the function of the protein. Different host cells have characteristic and specific mechanisms for the post-translational processing and modification of proteins. Appropriate cells lines or host systems can be chosen to ensure the correct modification and processing of the foreign protein expressed. To this end, eukaryotic host cells which possess the cellular machinery for proper processing of the primary transcript, glycosylation, and phosphorylation of the gene product may be used. Such mammalian host cells include but are not limited to CHO, VERO, BHK, HeLa, COS, MDCK, 293, WI38, PC12 etc.

Stable expression is preferred for long-term, high-yield production of recombinant proteins in animal cells. Rather than using expression vectors which contain viral origins of replication, host cells can be transformed with RPTP $\alpha$ , RPTP $\epsilon$ , or IR DNA controlled by appropriate expression control elements (e.g., promoter, enhancer, sequences, transcription terminators, polyadenylation sites, etc.), and a selectable marker. Following the introduction of

- 13 -

foreign DNA, engineered cells may be allowed to grow for 1-2 days in an enriched media, and then are switched to a selective media. The selectable marker  
5 in the recombinant plasmid confers resistance to the selection and allows cells to stably integrate the plasmid into their chromosomes and grow to form foci which in turn can be cloned and expanded into cell lines. This method, which is demonstrated in the  
10 examples below, may advantageously be used to engineer cell lines which stably coexpress both the RTP and IR-PTK, and which respond to ligand mediated signal transduction. Such engineered cell lines are particularly useful in screening PTP inhibitors,  
15 stimulators and analogs.

A number of selection systems may be used, including but not limited to the herpes simplex virus thymidine kinase (Wigler, et al., 1977, Cell 11:223), hypoxanthine-guanine phosphoribosyltransferase  
20 (Szybalska & Szybalski, 1962, Proc. Natl. Acad. Sci. USA 48:2026), and adenine phosphoribosyltransferase (Lowy, et al., 1980, Cell 22:817) genes can be employed in tk<sup>-</sup>, hgp<sup>r</sup>t<sup>-</sup> or ap<sup>r</sup>t<sup>-</sup> cells, respectively. Also, antimetabolite resistance can be used as the  
25 basis of selection for dhfr, which confers resistance to methotrexate (Wigler, et al., 1980, Natl. Acad. Sci. USA 77:3567; O'Hare, et al., 1981, Proc. Natl. Acad. Sci. USA 78:1527); gpt, which confers resistance to mycophenolic acid (Mulligan & Berg, 1981), Proc.  
30 Natl. Acad. Sci. USA 78:2072); neo, which confers resistance to the aminoglycoside G-418 (Colberre-Garapin, et al., 1981, J. Mol. Biol. 150:1); and hyg<sup>r</sup>, which confers resistance to hygromycin (Santerre, et al., 1984, Gene 30:147) genes.  
35 Recently, additional selectable genes have been described, namely trpB, which allows cells to utilize

- 14 -

indole in place of tryptophan; hisD, which all ws  
cells to utilize histinol in place of histidine.  
(Hartman & Mulligan, 1988, Proc. Natl. Acad. Sci. USA  
5 85:8047); and ODC (ornithine decarboxylase) which  
confers resistance to the ornithine decarboxylase  
inhibitor, 2-(difluoromethyl)-DL-ornithine, DFMO  
(McConlogue L., 1987, In: Current Communications in  
Molecular Biology, Cold Spring Harbor Laboratory ed.).

10 As the IR-PTK and RPTP may be coexpressed from  
different expression plasmids in the same cell, a  
different amplifiable selection system (for example,  
dhfr and adenosine deaminase) may be used for each  
individual plasmid. By applying different  
15 concentrations of the selecting drugs, the expression  
level of individual protein may be controlled  
separately as required (Wood et al., 1990, J. Immunol.  
145:3011-16).

The host cells which contain the coding sequences  
20 and which express the biologically active gene  
products may be identified by at least three general  
approaches; (a) DNA-DNA or DNA-RNA hybridization; (b)  
the presence or absence of "marker" gene functions;  
and (c) detection of the gene products as measured by  
25 immunoassay or by their biological activity.

In the first approach, the presence of the RPTP $\alpha$ ,  
RPTP $\epsilon$  or IR coding sequence(s) inserted in the  
expression vector(s) can be detected by DNA-DNA or  
DNA-RNA hybridization using probes comprising  
30 nucleotide sequences that are homologous to the RPTP $\alpha$ ,  
RPTP $\epsilon$  or IR coding sequence(s), respectively, or  
portions or derivatives thereof.

In the second approach, the recombinant expres-  
sion vector/host system can be identified and selected  
35 based upon the presence or absence of certain "marker"  
gene functions (e.g., thymidine kinase activity,



- 15 -

resistance to antibiotics, resistance to methotrexate, transformation phenotype, occlusion body formation in baculovirus, etc.). For example, if the RPTP $\alpha$ , RPTP $\epsilon$  or IR coding sequence(s) is inserted within a marker gene sequence of the vector, recombinant cells containing the RPTP $\alpha$ , RPTP $\epsilon$  or IR coding sequence(s) can be identified by the absence of the marker gene function. Alternatively, a marker gene can be placed in tandem with the RPTP $\alpha$ , RPTP $\epsilon$  or IR sequence under the control of the same or different promoter used to control the expression of the RPTP $\alpha$ , RPTP $\epsilon$  or IR coding sequence(s). Expression of the marker in response to induction or selection indicates expression of the RPTP $\alpha$ , RPTP $\epsilon$  or IR coding sequence(s).

In the third approach, the expression of the RPTP $\alpha$ , RPTP $\epsilon$  or IR protein product can be assessed immunologically, for example by Western blots, immunoassays such as immunoprecipitation, enzyme-linked immunoassays and the like. The ultimate test of the success of the expression system, however, involves the detection of the biologically active RPTP $\alpha$ , RPTP $\epsilon$  or IR proteins. A number of assays can be used to detect activity including but not limited to ligand binding assays, phosphorylation assays, dephosphorylation assays, and biological assays using engineered cell lines as the test substrate.

The RPTP $\alpha$ , RPTP $\epsilon$  or IR gene products as well as host cells or cell lines transfected or transformed with recombinant RPTP $\alpha$ , RPTP $\epsilon$  and IR expression vector(s) can be used for a variety of purposes. These include but are not limited to the screening and selection of RPTP $\alpha$  or RPTP $\epsilon$  analogs, or drugs that act by interacting with RTP-IR-PTK complex, or generating antibodies (i.e., monoclonal or polyclonal) that bind

- 16 -

to the RTP-IR-PTK complex, including those that competitively inhibit the formation of such complexes. These gene products or host cells or cell lines may  
5 also be used for identifying other signalling molecules or their genes that are engaged in the insulin signalling pathway.

## 5.2 ASSAY SYSTEMS FOR DRUG SCREENING

10

In one embodiment of the invention, the RPTPs, the RTP-IR-PTK complex, or cell lines that express the RPTPs or RTP-IR-PTK complex, may be used to screen for molecules that modulate RTP activity.

15

Such molecules may include small organic or inorganic compounds, antibodies, peptides, or other molecules that modulate RTP $\alpha$ 's or RTP $\epsilon$ 's dephosphorylation activity toward IR, or that promote or prevent the formation of RTP $\alpha$ -IR or RTP $\epsilon$ -IR complex. Synthetic  
20 compounds, natural products, and other sources of potentially biologically active materials can be screened in a number of ways.

25

The ability of a test molecule to modulate the activity of RTP $\alpha$  or RTP $\epsilon$  toward IR, hence signal transduction, may be measured using standard biochemical techniques, such as those described in section 6.1. Other responses such as activation or suppression of catalytic activity, phosphorylation or dephosphorylation of other proteins, activation or  
30 modulation of second messenger production, changes in cellular ion levels, association, dissociation or translocation of signalling molecules, or transcription or translation of specific genes may also be monitored. These assays may be performed  
35 using conventional techniques developed for these purposes in the course of screening.

- 17 -

Ligand binding to its cellular receptor may, via signal transduction pathways, affect a variety of cellular processes. Cellular processes under the control of insulin signalling pathway may include, but are not limited to, normal cellular functions such as carbohydrate metabolism, proliferation, differentiation, maintenance of cell shape, and adhesion, in addition to abnormal or potentially deleterious processes such as apoptosis, loss of contact inhibition, blocking of differentiation or cell death. The qualitative or quantitative observation and measurement of any of the described cellular processes by techniques known in the art may be advantageously used as a means of scoring for signal transduction in the course of screening.

Described in this section are methods of the invention for screening, identification and evaluation of compounds that interact with RPTP $\alpha$ , RPTP $\epsilon$  and IR and may affect various cellular processes under the control of the insulin signalling pathway.

The present invention includes a method for identifying a compound which is capable of, by modulating phosphotyrosine phosphatase activity of RPTP $\alpha$  and/or RPTP $\epsilon$ , modulating insulin receptor type protein kinase IR-PTK signal transduction, comprising:

- (a) contacting the compound with RPTP $\alpha$  and/or RPTP $\epsilon$  and IR or, a functional derivatives thereof, in pure form, in a membrane preparation, or in a whole live or fixed cell;
- (b) incubating the mixture of step (a) for an interval sufficient for the compound to stimulate or inhibit the phosphotyrosine phosphatase enzymatic activity or the signal transduction;

- 18 -

- (c) measuring the phosphotyrosine phosphatase enzymatic activity or the signal transduction;
- 5 (d) comparing the phosphotyrosine phosphatase enzymatic activity or the signal transduction activity to that of RPTP $\alpha$ , and/or RPTP $\epsilon$  and IR, incubated without the compound, thereby determining whether the
- 10 compound stimulates or inhibits signal transduction.

RPTP $\alpha$  and/or RPTP $\epsilon$  and IR, or functional derivatives thereof, for example, having amino acid deletions and/or insertions and/or substitutions while

15 maintaining signal transduction, can also be used for the testing of compounds. A functional derivative may be prepared from a naturally occurring or recombinantly expressed RPTP $\alpha$ , RPTP $\epsilon$  and IR by proteolytic cleavage followed by conventional

20 purification procedures known to those skilled in the art. Alternatively, the functional derivative may be produced by recombinant DNA technology by expressing only these parts of RPTP $\alpha$ , RPTP $\epsilon$  or IR in suitable cells. Cells expressing RPTP $\alpha$  and/or RPTP $\epsilon$  and IR may

25 be used as a source of RPTP $\alpha$ , RPTP $\epsilon$  and/or IR, crude or purified, or in a membrane preparation, for testing in these assays. Alternatively, whole live or fixed cells may be used directly in those assays. The cells may be genetically engineered to coexpress RPTP $\alpha$ ,

30 RPTP $\epsilon$  and IR. The cells may also be used as host cells for the expression of other recombinant molecules with the purpose of bringing these molecules into contact with RPTP $\alpha$ , RPTP $\epsilon$  and/or IR within the cell.

35 IR-PTK signal transduction activity may be measured by standard biochemical techniques or by

- 19 -

monitoring the cellular processes controlled by the signal. To assess modulation of phosphatase activity, the test molecule is added to a reaction mixture  
5 containing the phosphorylated substrate and the phosphatase. To assess modulation of kinase activity of the IR-PTK, the test molecule is added to a reaction mixture containing the IR-PTK and its substrate (in the case of autophosphorylation, the IR-  
10 PTK is also the substrate). Where the test molecule is intended to mimic ligand stimulation the assay is conducted in the absence of insulin. Where the test molecule is intended to reduce or inhibit insulin activity the presence of insulin. The kinase reaction  
15 is then initiated with the addition of ATP. An immunoassay is performed on the kinase or phosphatase reaction to detect the presence or absence of the phosphorylated tyrosine residues on the substrate, and results are compared to those obtained for controls  
20 i.e., reaction mixtures not exposed to the test molecule. The immunoassay used to detect the phosphorylated substrate in the cell lysate or the *in vitro* reaction mixture may be carried out with an anti-phosphotyrosine antibody. Signal transduction is  
25 mimicked if the cellular processes under the control of the signalling pathway are affected in a way similar to that caused by ligand binding. Such compounds may be naturally occurring or synthetically produced molecules that could replace the  
30 administration of insulin in the treatment of diabetes.

The invention also includes a method whereby a molecule capable of binding to RPTP $\alpha$  and/or RPTP $\epsilon$  and IR in a chemical or biological preparation may be  
35 identified comprising:

- 20 -

- (a) immobilizing RPTP $\alpha$  and/ or RPTP $\epsilon$  and IR, or fragments thereof, to a solid phase matrix;
- (b) contacting the chemical or biological preparation with the solid phase matrix produced in step (a), for an interval sufficient to allow the compound to bind;
- (c) washing away any unbound material from the solid phase matrix;
- (d) detecting the presence of the compound bound to the solid phase,

thereby identifying the compound.

The above method may further include the step of:

- (e) eluting the bound compound from the solid phase matrix, thereby isolating the compound.

The term "compound capable of binding to RPTP $\alpha$  and/or RPTP $\epsilon$  and IR" refers to a naturally occurring or synthetically produced molecule which interacts with RPTP $\alpha$  and/or RPTP $\epsilon$  and IR. Such a compound may directly or indirectly modulate IR-PTK signal transduction and may include molecules that are natively associated with RPTP $\alpha$ , RPTP $\epsilon$  and/or IR inside a cell. Examples of such compounds are (i) a natural substrate, of RPTP $\alpha$  and/or RPTP $\epsilon$ ; (ii) a naturally occurring molecule which is part of the signalling complex; iii) a natural substrate of IR-PTK, iv) a naturally occurring signalling molecule produced by other cell types.

The present invention also includes methods for identifying the specific site(s) of RPTP $\alpha$ , or RPTP $\epsilon$  interaction with IR. Using the methods described herein, and biochemical and molecular biological methods well-known in the art, it is possible to identify the corresponding portions of RPTP $\alpha$ , RPTP $\epsilon$  and IR involved in this interaction. For example,

site-directed mutagenesis of DNA encoding either RPTP $\alpha$ , RPTP $\epsilon$  or IR may be used to destroy or inhibit the interaction between the two molecules.

5 Biophysical methods such as X-ray crystallography and nuclear magnetic resonance may also be used to map and study these sites of interaction. Once these sites have been identified, the present invention provides means for promoting or inhibiting this interaction,  
10 depending upon the desired biological outcome. Based on the foregoing, given the physical information on the sites of interaction is known, compounds that modulate catalytic activity and signal transduction may be elaborated by standard methods well known in  
15 the field of rational drug design.

The present invention further provides an assay for identifying a compound, which can block the interaction of RPTP $\alpha$  or RPTP $\epsilon$  and IR. For example, a cell transfected to coexpress RPTP $\alpha$  or RPTP $\epsilon$  and IR,  
20 in which the two proteins interact to form a RPTP $\alpha$ -IR or RPTP $\epsilon$ -IR complex, can be incubated with an agent suspected of being able to inhibit this interaction, and the effect on the interaction measured. Any of a number of means for measuring the interaction and its  
25 disruption, such as coimmunoprecipitation, are available. The present invention also provides an assay method to identify and test a compound which stabilizes and promotes the interaction, using the same approach described above for a potential  
30 inhibitor.

Random peptide libraries consisting of all possible combinations of amino acids may be used to identify peptides that are able to bind to the substrate binding site of RPTP $\alpha$  or RPTP $\epsilon$ , or other  
35 functional domains of RPTP $\alpha$  or RPTP $\epsilon$ . Similarly, such libraries may also be used to identify peptides

- 22 -

that are able to bind to the IR's site of interaction with RPTP $\alpha$  or RPTP $\epsilon$ . Identification of molecules that are able to bind to RPTP $\alpha$ , RPTP $\epsilon$  and IR may be accomplished by screening a peptide library with recombinant RPTP $\alpha$ , RPTP $\epsilon$  or IR proteins or recombinant soluble forms of RPTP $\alpha$  or RPTP $\epsilon$  or IR protein. Alternatively, the phosphatase and extracellular ligand binding domains of RPTP $\alpha$  or RPTP $\epsilon$  may be separately expressed and used to screen peptide libraries.

One way to identify and isolate the peptide that interacts and forms a complex with RPTP $\alpha$  or RPTP $\epsilon$  and IR, may involve labeling or "tagging" RPTP $\alpha$  or RPTP $\epsilon$  and IR proteins. The RPTP $\alpha$  or RPTP $\epsilon$  and IR proteins may be conjugated to enzymes such as alkaline phosphatase or horseradish peroxidase or to other reagents such as fluorescent labels which may include fluorescein isothiocyanate (FITC), phycoerythrin (PE) or rhodamine. Conjugation of any given label, to RPTP $\alpha$  or RPTP $\epsilon$  and IR, may be performed using techniques that are routine in the art. Alternatively, RPTP $\alpha$ , RPTP $\epsilon$  or IR expression vectors may be engineered to express a chimeric RPTP $\alpha$ , RPTP $\epsilon$  or IR protein containing an epitope for which a commercially available antibody exists. The epitope specific antibody may be tagged using methods well known in the art including labeling with enzymes, fluorescent dyes or colored or magnetic beads.

The present invention also includes a method for identifying and isolating a nucleic acid molecule encoding a gene product which is capable of, by modulating phosphotyrosine phosphatase activity RPTP $\alpha$  and/or RPTP $\epsilon$ , modulating IR-PTK signal transduction, comprising:



- 23 -

- (a) introducing the nucleic acid molecule into host cells coexpressing RPTP $\alpha$  and/or RPTP $\epsilon$  and IR or fragments thereof;
- 5 (b) culturing the cells so that the gene product encoded by the nucleic acid molecule is expressed in the host cells and interacts with RPTP $\alpha$  and/or RPTP $\epsilon$  and IR or fragments thereof;
- 10 (c) measuring the phosphotyrosine phosphatase enzymatic activity of RPTP $\alpha$  and/or RPTP $\epsilon$  or IR-PTK signal transduction activity;
- (d) comparing the phosphotyrosine phosphatase enzymatic activity or signal transduction to that of RPTP $\alpha$  and/or RPTP $\epsilon$  and IR, or fragments thereof in cells without the nucleic acid molecule, thereby determining whether the gene product encoded by the nucleic acid molecule modulates IR-PTK signal transduction.
- 15 20

The above method may further include the step of:

- (e) selecting and culturing the cells identified in step (d), recovering the nucleic acid molecule, thereby isolating the nucleic acid molecule.
- 25

By the term "nucleic acid molecule" is meant a naturally occurring or recombinantly generated nucleic acid molecule containing a nucleotide sequence operatively associated with an element that controls expression of the nucleotide sequence. An expression library may be created by introducing into host cells a pool of different nucleic acid molecules encoding different gene products. The host cells may be genetically engineered to coexpress RPTP $\alpha$ , RPTP $\epsilon$  and IR. Such a gene library may be screened by standard biochemical techniques or by monitoring the cellular

30 35

- 24 -

processes controlled by the signal. This approach is especially useful in identifying other native signalling molecules that are also involved in the signalling pathway.

### 5.3 ANTIBODY PRODUCTION AND SCREENING

Various procedures known in the art may be used for the production of antibodies to epitopes of the recombinantly produced RPTP $\alpha$ , RPTP $\epsilon$ , IR, RPTP $\alpha$ -IR and RPTP $\epsilon$ -IR complex. Such antibodies include but are not limited to polyclonal, monoclonal, chimeric, single chain, Fab fragments and fragments produced by an Fab expression library. Neutralizing antibodies *i.e.*, those which compete for the substrate binding site of RPTP $\alpha$  or RPTP $\epsilon$ , or the IR's site of interaction with RPTP $\alpha$  or RPTP $\epsilon$  are especially preferred for therapeutics.

For the production of antibodies, various host animals may be immunized by injection with RPTP $\alpha$ , RPTP $\epsilon$ , IR, RPTP $\alpha$ -IR or RPTP $\epsilon$ -IR complex, or genetically engineered cells expressing RPTP $\alpha$ , RPTP $\epsilon$  and IR, including but not limited to rabbits, mice, rats, etc. Various adjuvants may be used to increase the immunological response, depending on the host species, including but not limited to Freund's (complete and incomplete), mineral gels such as aluminum hydroxide, surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, keyhole limpet hemocyanin, dinitrophenol, and potentially useful human adjuvants such as BCG (bacille Calmette-Guerin) and Corynebacterium parvum.

Monoclonal antibodies to RPTP $\alpha$ , RPTP $\epsilon$ , IR, RPTP $\alpha$ -IR and RPTP $\epsilon$ -IR complex may be prepared by using any

- 25 -

technique which provides for the production of antibody molecules by continuous cell lines in culture. These include but are not limited to the hybridoma technique originally described by Kohler and Milstein, (Nature, 1975, 256:495-497), the human B-cell hybridoma technique (Kosbor et al., 1983, Immunology Today, 4:72; Cote et al., 1983, Proc. Natl. Acad. Sci., 80:2026-2030) and the EBV-hybridoma technique (Cole et al., 1985, Monoclonal Antibodies and Cancer Therapy, Alan R. Liss, Inc., pp. 77-96). In addition, techniques developed for the production of "chimeric antibodies" (Morrison et al., 1984, Proc. Natl. Acad. Sci., 81:6851-6855; Neuberger et al., 1984, Nature, 312:604-608; Takeda et al., 1985, Nature, 314:452-454) by splicing the genes from a mouse antibody molecule of appropriate antigen specificity together with genes from a human antibody molecule of appropriate biological activity can be used. Alternatively, techniques described for the production of single chain antibodies (U.S. Patent 4,946,778) can be adapted to produce RPTP $\alpha$ , RPTP $\epsilon$ , IR, RPTP $\alpha$ -IR or RPTP $\epsilon$ -IR complex-specific single chain antibodies.

Antibody fragments which contain specific binding sites of RPTP $\alpha$ , RPTP $\epsilon$ , IR, RPTP $\alpha$ -IR or RPTP $\epsilon$ -IR complex may be generated by known techniques. For example, such fragments include but are not limited to: the F(ab')<sub>2</sub> fragments which can be produced by pepsin digestion of the antibody molecule and the Fab fragments which can be generated by reducing the disulfide bridges of the F(ab')<sub>2</sub> fragments. Alternatively, Fab expression libraries may be constructed (Huse et al., 1989, Science, 246:1275-1281) to allow rapid and easy identification of monoclonal Fab

- 26 -

fragments with the desired specificity to RPTP $\alpha$ , RPTP $\epsilon$ , IR, RPTP $\alpha$ -IR or RPTP $\epsilon$ -IR complex.

5                    6.    EXAMPLE: DEMONSTRATION OF AN IN VIVO  
                         SELECTION SYSTEM FOR INSULIN RECEPTOR  
                         ACTIVATION

                         In the example described below, host cells were  
                         engineered to express both the IR and a series of  
10    PTPs. The cells expressing IR alone or IR plus an  
                         ineffective PTP display an altered phenotype when  
                         exposed to insulin. The results show that  
                         coexpression of RPTP $\alpha$  or RPTP $\epsilon$  inhibits  
                         phosphorylation of the IR and restores normal cell  
15    phenotype. The results demonstrate that RPTP- $\alpha$  and  
                         RPTP- $\epsilon$  modulate with IR signal transduction.

                         6.1    MATERIAL AND METHODS

20                    IR/BHK cells were maintained in DMEM/high  
                         glucose, 10% fetal calf serum, 10 mM glutamine, 1  $\mu$ M  
                         methotrexate plus antibiotics. The cDNAs for RPTP $\alpha$  or  
                         RPTP $\epsilon$  were cloned into a cytomegalovirus early  
                         promoter-based expression plasmid pCMV (Eaton et al.,  
25    Biochemistry, 25:8343-47, 1986). Plasmid DNA were  
                         transfected into 10<sup>7</sup> BHK cells/10cm<sup>2</sup> plate according to  
                         the protocol of Chen and Okayama (Mol. Cell Biol.,  
                         7:2745-52, 1987). Eighteen hours after the addition  
                         of DNA precipitate, cells were washed once and  
30    supplied with fresh medium containing 0.5% serum.  
                         Forty-eight hours after transfection, the cells were  
                         split at least 1:10. Medium containing 1  $\mu$ M insulin  
                         was added 12 hours later. Medium containing insulin  
                         was changed 3 times a day. Cells in culture were  
35    washed thoroughly with PBS each time the media was  
                         changed in order to remove detached cells.

- 27 -

The presence of insulin does not cause cell death, but detachment, so it is necessary to maintain the selective pressure of insulin presence until stable co-transfected clones have grown to sufficient numbers to be isolated and characterized. This process took approximately four weeks.

Antibodies used in the analysis of protein expression and phosphorylation were the mouse monoclonal antiphosphotyrosine antibody 5E2 (Fendly et al., 1990, Cancer Res., 50:1550-8), mouse anti-IR monoclonal antibody 18-34 and rabbit antisera against the phosphatases. The rabbit antisera to RPTP $\alpha$  and RPTP $\epsilon$  were prepared by standard techniques using peptide fragments derived from the C-terminus of RPTP $\alpha$  and RPTP $\epsilon$  as immunogen. For detection of phosphotyrosine and protein antigens on immunoblots, the ECL system (Amersham) was used in conjunction with goat anti-mouse and anti-rabbit antibodies (Biorad). For reprobing, blots were stripped in 67 mM Tris-HCl (pH 6.8), 2% SDS, and 0.1%  $\beta$ -mercaptoethanol at 50°C for 30 minutes.

## 6.2 SELECTION AND ANALYSIS OF CELLS BY TRANSFECTION WITH cDNAS ENCODING PTPS

The specificity of each PTP for the insulin receptor was determined by assaying insulin-induced phenotypic changes in the cells and phosphorylation of insulin receptor  $\beta$ -subunit by Western blot as described below.

### 6.2.1 INSULIN-INDUCED CHANGE IN PHENOTYPE

In the presence of 1  $\mu$ M insulin IR/BHK cells display an abnormal phenotype, i.e., rounding up and becoming detached from the plastic surface (Figure

- 28 -

1A). The change in the morphology and the loss of adhesion to the substratum induced by insulin was most pronounced at low cell density and in the presence of 10% fetal calf serum. IR/BHK cells were transfected with cDNAs coding for PTP1B, PTP1BΔ299, PTP1C, PTPμ, CD45, RPTPκ, RPTPα, RPTPε, LAR, and LAR(domain 1). To determine which of these PTPs were capable of modulating IR activity thereby preventing these phenotypic changes of the cells. Only RPTPα and RPTPε, were able to restore the normal phenotype. After 24 hours of selection, small clones consisting of 4-8 cells could be seen. These transfected cells exhibited the normal phenotype and did not respond in the same manner to high doses of insulin as the cells transfected with IR alone (Figure 3B).

#### 6.2.2 AUTOPHOSPHORYLATION ASSAY BY WESTERN BLOT

Two stably cotransfected clones for each transfection (IR + RPTPα and IR + RPTPε) were starved overnight in DMEM/high glucose containing 0% fetal calf serum then stimulated with 1 μM insulin for 10 minutes. The cells were lysed and the phosphotyrosine content of insulin receptor β-subunit was detected by Western blotting (Figures 2 and 3) using anti-phosphotyrosine antibodies.

Figure 2A shows the phosphorylation status of IR in stable BHK cell clones coexpressing IR and RPTPα. In control cells a strong tyrosine phosphorylation of insulin receptors β-subunit could be detected. This phosphorylation level was lower with the clones obtained after transfection with cDNA encoding RPTPα. Figure 2B shows the level of RPTPα expression in the cotransfected clones. An additional band immunoreactive with anti-RPTPα antibodies, could be

- 29 -

detected in these cotransfected clones. Figure 2C shows the level of IR expression in control and cotransfected clones, which was similar. Stable BHK  
5 cell clone 5 coexpressing IR and RPTP $\alpha$  was deposited with the American Type Culture Collection, 12301 Parklawn Drive, Rockville, MD 20852 on January 20, 1994, and assigned accession number ATCC CRL 11528.

As shown in Figure 3A, 3B and 3C, the pattern of  
10 IR phosphorylation and protein expression levels in stable cell clones coexpressing IR and RPTP $\epsilon$  are similar to that of IR and RPTP $\alpha$ . The data suggests that the restoration of normal phenotype of the cotransfected cells was associated with the  
15 dephosphorylation of the insulin receptor or downstream key signalling event. Stable BHK cell clone 6 coexpressing IR and RPTP $\epsilon$  was deposited with the American Type Culture Collection, 12301 Parklawn Drive, Rockville, MD 20852 on January 20, 1994, and  
20 assigned accession number ATCC CRL 11529.

The results described clearly indicate that RPTP $\alpha$  and RPTP $\epsilon$  interact specifically with IR. In the presence of insulin, RPTP $\alpha$  and RPTP $\epsilon$  modulate IR signal transduction and downstream cellular processes,  
25 which prevent changes in cell morphology and adhesion properties. These cell lines could be used in a drug screen whereby any biological effect of the test compound *in vivo* on insulin signal transduction may be monitored by changes in the cell morphology and  
30 adhesion properties or by phosphorylation state of the insulin receptor. Drugs that interfere with RPTP $\alpha$  or RPTP $\epsilon$  activity would make the cells respond to insulin and re-exhibit the insulin-sensitive phenotype.

The present invention is not to be limited in  
35 scope by the specific embodiments described which are intended as single illustrations of individual aspects

- 30 -

of the invention, and functionally equivalent methods  
and components are within the scope of the invention.  
Indeed, various modifications of the invention, in  
5 addition to those shown and described herein will  
become apparent to those skilled in the art from the  
foregoing description and accompanying drawings. Such  
modifications are intended to fall within the scope of  
the appended claims.

10

15

20

25

30

35



- 31 -

## WHAT IS CLAIMED IS:

1. A genetically engineered mammalian cell  
5 containing:
  - (a) a first nucleic acid molecule having a nucleotide sequence which encodes a PTP or a fragment thereof, operatively associated with an element that controls its  
10 expression; and
  - (b) a second nucleic acid molecule having a nucleotide sequence which encodes a IR-PTK or a fragment thereof, operatively associated with an element that controls its  
15 expression;whereby a PTP and a IR-PTK are coexpressed by the mammalian cell.
2. The cell of Claim 1 in which the IR-PTK is  
20 human IR.
3. The cell of Claim 2 in which the PTP is human RPTP $\alpha$ .
- 25 4. The cell of Claim 2 in which the PTP is human RPTP $\epsilon$ .
5. The cell of Claim 3 in which the cell is a stably transfected baby hamster kidney cell.  
30
6. The cell of Claim 4 in which the cell is a stably transfected baby hamster kidney cell.
7. The cell of Claim 5 as deposited with the  
35 American Type Culture Collection, and assigned accession number ATCC CRL 11528.

- 32 -

8. The cell of Claim 6 as deposited with the American Type Culture Collection and assigned accession number ATCC CRL 11529.

5

9. A method for determining whether a compound is capable of, by modulating phosphotyrosine phosphatase activity of RPTP $\alpha$  or RPTP $\epsilon$ , modulating IR-PTK signal transduction, comprising:

10

(a) contacting the compound with a whole live or fixed cell of Claim 1, for an interval sufficient for the compound to modulate the signal transduction;

(b) measuring the signal transduction; and

15

(c) comparing the signal transduction to that incubated without the compound,

thereby determining whether the compound modulates the signal transduction.

20

10. A method for identifying a nucleic acid molecule encoding a gene product which is capable of modulating IR-PTK signal transduction by modulating the enzymatic activity of phosphotyrosine phosphatase, comprising:

25

(a) introducing the nucleic acid molecule into the cells of Claim 1;

(b) culturing the cells so that the gene product encoded by the nucleic acid molecule is expressed in the cells and interacts with the phosphotyrosine phosphatase and IR-PTK or functional derivatives thereof;

30

(c) measuring the signal transduction; and

(d) comparing the signal transduction to that in the cells without the nucleic acid molecule

35

thereby determining whether the gene product encoded by the nucleic acid molecule is capable of modulating the signal transduction.

5

11. A method for isolating from a mixture the nucleic acid molecule of Claim 10, comprising the steps (a) through (d) of claim 10 and:

10 (e) selecting and culturing the cells identified in step (d), recovering the nucleic acid molecule, thereby isolating the nucleic acid molecule.

12. The method of Claim 10 in which the signal  
15 transducing activity is stimulated.

13. The method of Claim 11 in which the signal transducing activity is stimulated.

20

25

30

35

1/5



FIG. 1A

SUBSTITUTE SHEET (RULE 26)

2/5

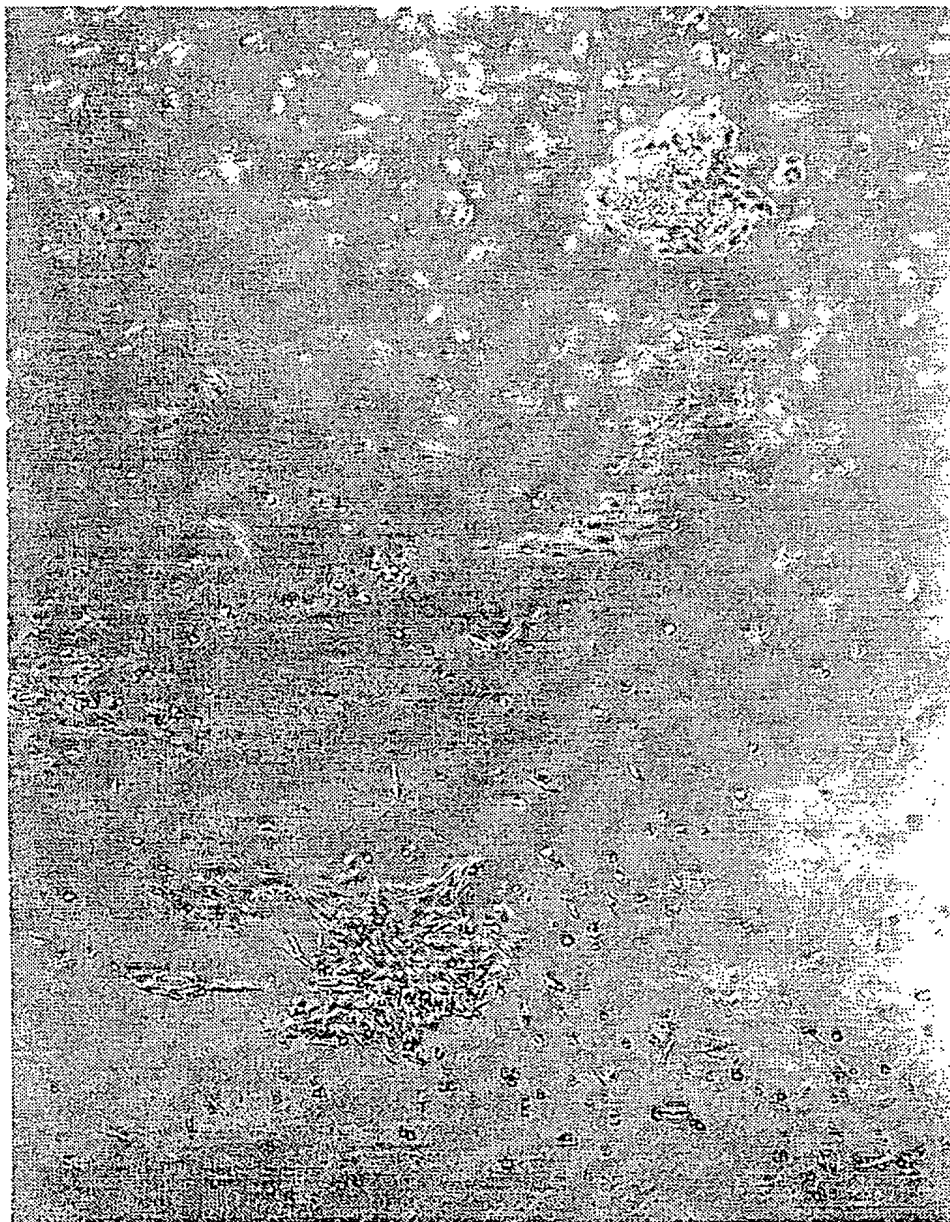


FIG. 1B

3/5

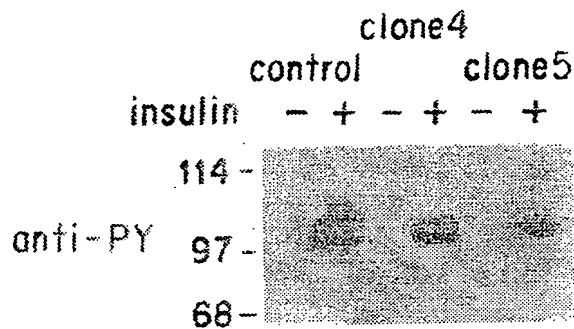


FIG.2A

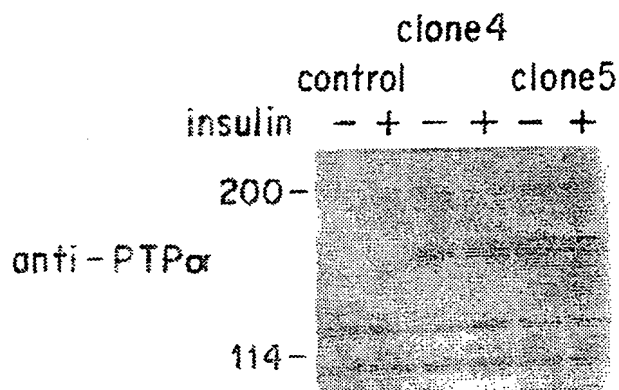


FIG.2B

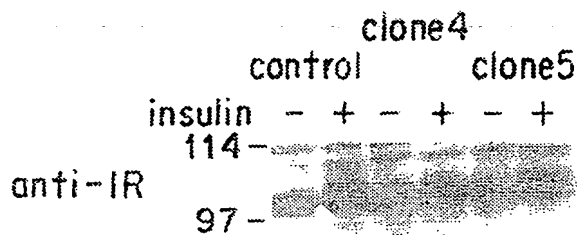


FIG.2C

4/5

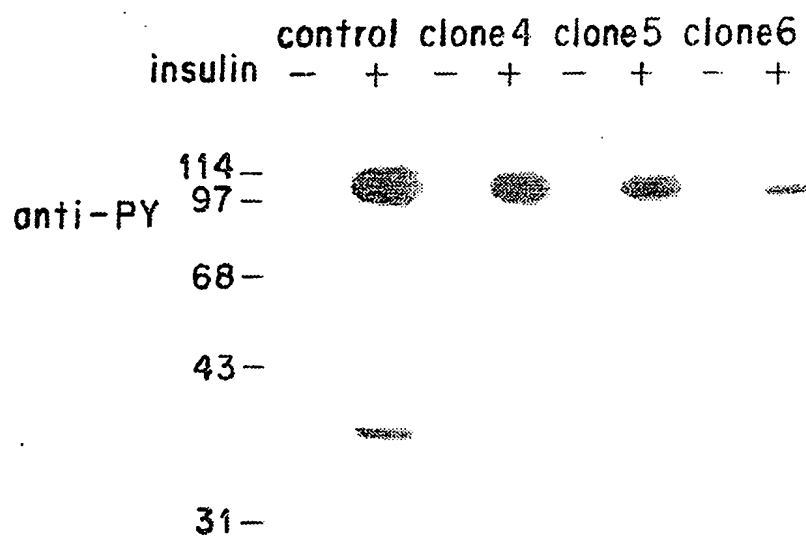


FIG.3A

5/5

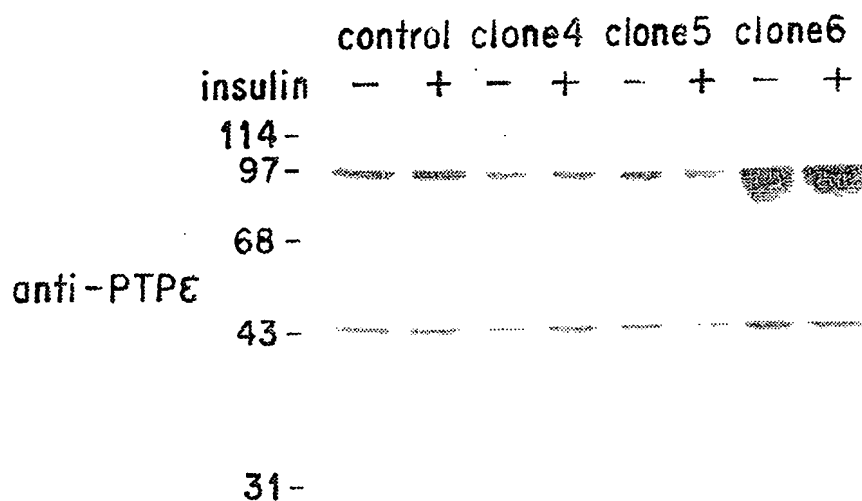


FIG.3B

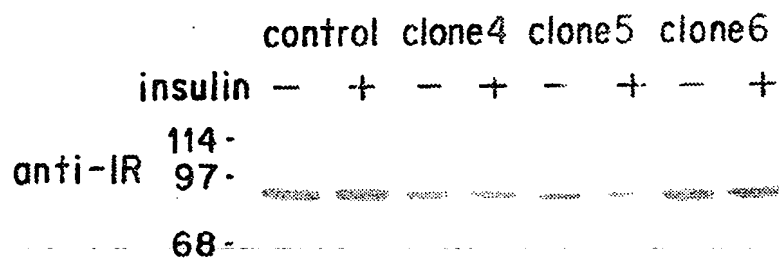


FIG.3C



# INTERNATIONAL SEARCH REPORT

International Application No  
PCT/EP 95/00731

## A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/85 C12N5/10 C12Q1/42 C12Q1/48 C07K14/72  
//C12N9/16

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N C12Q

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	J CELL BIOL 116 (3). 1992. 627-634. CODEN: JCLBA3 ISSN: 0021-9525, TARTARE S ET AL 'ACTIVATION OF INSULIN-EPIDERMAL GROWTH FACTOR EGF RECEPTOR CHIMERAE REGULATES EGF RECEPTOR BINDING AFFINITY.' see page 630, column 1, paragraph 3 ---	1-6
A	US-A-5 155 031 (POSNER ET AL) 13 October 1992 cited in the application ---	
A	BIOCHEM BIOPHYS RES COMMUN 178 (3). 1991. 1291-1297. CODEN: BBRC99 ISSN: 0006-291X, ZHANG W-R ET AL 'IDENTIFICATION OF SKELETAL MUSCLE PROTEIN-TYROSINE PHOSPHATASES BY AMPLIFICATION OF CONSERVED COMPLEMENTARY DNA SEQUENCES.' ---	
-/--		

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

### \* Special categories of cited documents :

- \*A\* document defining the general state of the art which is not considered to be of particular relevance
- \*E\* earlier document but published on or after the international filing date
- \*L\* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- \*O\* document referring to an oral disclosure, use, exhibition or other means
- \*P\* document published prior to the international filing date but later than the priority date claimed

- \*T\* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- \*X\* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- \*Y\* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- \* & \* document member of the same patent family

Date of the actual completion of the international search

3 July 1995

Date of mailing of the international search report

5. 07. 95

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2  
NL - 2280 HV Rijswijk  
Tel. (+ 31-70) 340-2040, Tx. 31 651 epo nl,  
Fax (+ 31-70) 340-3016

Authorized officer

Molina Galan, E

# INTERNATIONAL SEARCH REPORT

Application No  
PCT/EP 95/00731

## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	J BIOL CHEM 268 (9). 1993. 6622-6628, YANG Q; CO D; SOMMERCORN J; TONKS N K 'CLONING AND EXPRESSION OF PTP-PEST A NOVEL HUMAN NONTRANSMEMBRANE PROTEIN TYROSINE PHOSPHATASE' ---	1
A	EP-A-0 245 979 (MERCK AND CO.) 19 November 1987 see page 2, line 22 - page 3, line 10 ---	
A	NATURE, vol. 313, 1985 LONDON GB, pages 756-761, ULLRICH ET AL. 'Human IR ...' cited in the application ---	
A	EMBO JOURNAL, vol. 9, no. 10, 1990 EYNHAM, OXFORD GB, pages 3241-3252, KRUEGER ET AL. 'Structural diversity ...' cited in the application ---	
A	PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA, vol. 87, 1990 WASHINGTON US, pages 6112-6116, SAP ET AL. 'Cloning and expression ...' cited in the application -----	

# INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/EP 95/00731

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
US-A-5155031	13-10-92	CA-A- 2042556	08-12-91
EP-A-245979	19-11-87	US-A- 4710469	01-12-87
		JP-A- 63022186	29-01-88

**THIS PAGE BLANK (USPTO)**